

COVID-19 Research Update



Andrzej Joachimiak

**Argonne National Laboratory
Structural Biology Center, X-ray Science Division, Advanced Photon Source
The University of Chicago
Center for Structural Genomics of Infectious Diseases**

October 23, 2020



Funding is provided by: the National Institute of Allergy and Infectious Diseases of NIH under Contract No. HHSN272201700060C, the DOE Office of Science through the National Virtual Biotechnology Laboratory, a consortium of DOE national laboratories focused on response to COVID-19 (the Coronavirus CARES Act) and the use of Structural Biology Center beamlines at the Advanced Photon Source is supported by the U.S. DOE Office of Science and operated for the DOE Office of Science by Argonne National Laboratory under Contract No. DE-AC02-06CH11357.

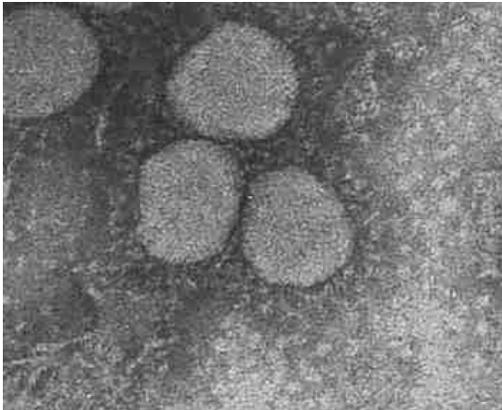


Argonne National Laboratory is a U.S. Department of Energy laboratory managed by UChicago Argonne, LLC.

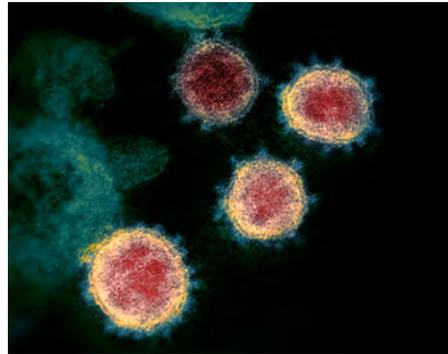
COVID-19 Pandemic : Images of SARS-CoV-2

- Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) is an etiologic agent responsible for the current outbreak of Coronavirus Disease 2019 (COVID-19) - at present there is no effective vaccine or proven drug to prevent infections and stop virus proliferation.

- SARS-CoV-2 isolated in FRhK-4 cells
- Thin section electron micrograph and negative stained virus particles

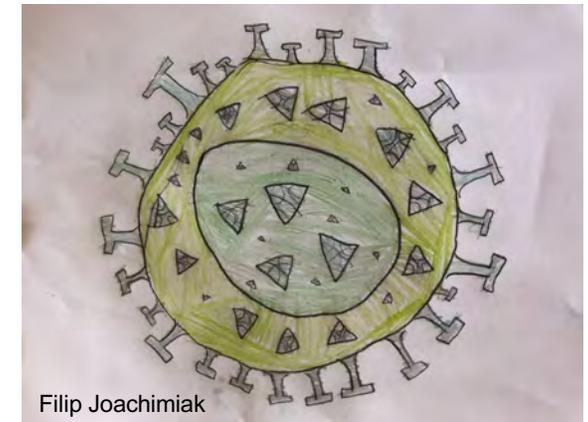
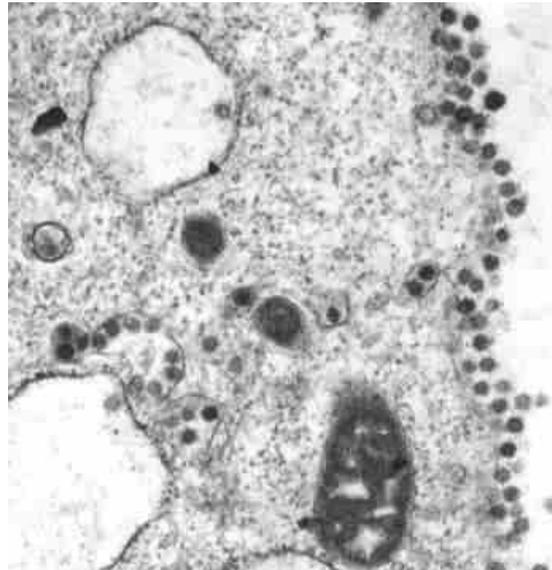


- Department of Microbiology, The University of Hong Kong and the Government Virus Unit, Department of Health, Hong Kong SAR China



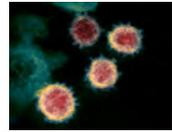
Infected: >41.5M
Death: >1.14 M

SARS-CoV-2 isolated from first US patient (NIAID)

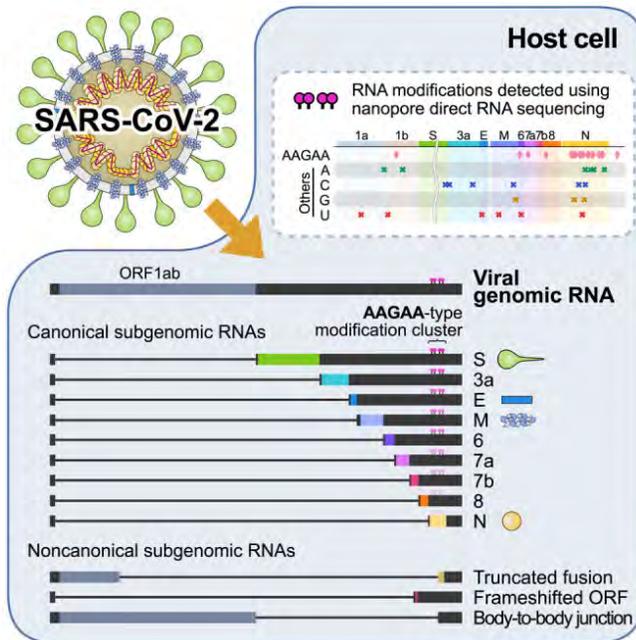
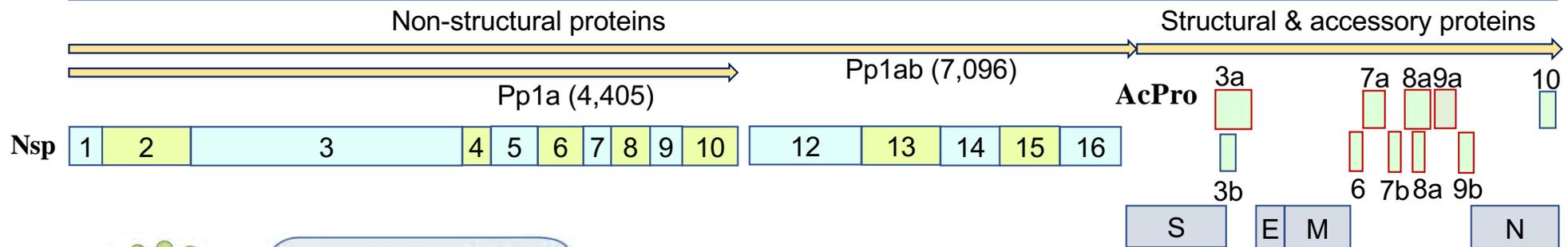


Filip Joachimiak

SARS-CoV-2 Genome



(+) RNA (29,903)



Kim *et al.* *Cell*, 2020

- SARS-CoV-2 is spherical, enveloped, non-segmented, (+) sense RNA betacoronavirus with a large ~30 kbs ssRNA genome.
- The RNA genome is coding for 29 proteins.
- The 4 structural, 15 non-structural and 9-10 accessory proteins are translated from subgenomic RNAs.
- Polyprotein processing is essential for the release and maturation of the 15 Nsps and assembly into cytoplasmic, ER membrane-bound multicomponent replicase-transcriptase complex.
- This complex is responsible for directing the replication, transcription and maturation of the viral genome and subgenomic mRNAs.
- Although this virus is similar to human and animal SARS- and MERS-CoVs, the detailed information about SARS-CoV-2 proteins structures and functions is urgently needed to rapidly develop effective therapeutics.

Center for Structural Genomics of Infectious Diseases



NIH: NIAID

We are a consortium of laboratories using state-of-the-art structural biology methods to determine the 3-D structures of proteins from pathogens in the NIAID Category A-C priority lists and organisms causing emerging and re-emerging infectious diseases. We do this as a free service to the scientific community! Members of the scientific community are encouraged to submit their targets of interest to CSGID by using our online form: [Submit/check your proposal now!](#)

For more detailed information about our mission, please refer to the [CSGID brochure \(pdf\)](#) and the [Overview](#).



Who we are?



Community Services



Scoreboard

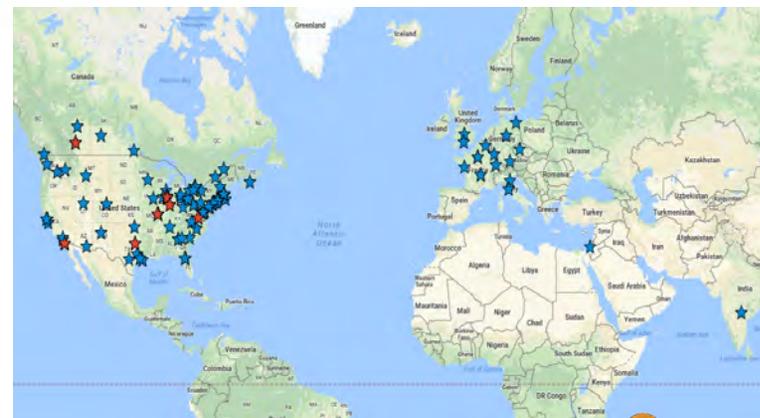


Structures



Publications

Targets:	12005
Targets requested by the community:	5391
Community Interest targets:	157
Targets with structures:	864
Structures:	1330 (X-ray: 1318, NMR: 7, Cryo-EM: 5)
Structures complexed with ligands:	560
Solved proteins requested by community:	385 (567 structures)
Solved Community Interest targets:	53 (122 structures)
Publications:	201 (131 structural, 70 methodological/other)
Citations:	4990 (2376 for structural, 2614 for methodological/other)



DOE broad capabilities for addressing COVID-19 crisis

- Light and neutron sources
- Nanoscience centers
- Computational resources
- People with deep expertise relevant to:
 - Testing
 - Antiviral drug discovery
 - Vaccine discovery
 - Supply chain bottlenecks
 - Modeling and understanding disease spread
 - Molecular and structural biology

HOW DOE AND OUR LABS ARE COMBATING COVID-19

UNDERSTANDING THE STRUCTURE –
DOE scientists are studying the components of the virus so we can determine how to fight it.

MODELING EPIDEMICS –
DOE scientists use previous experience they gained modeling Smallpox, Anthrax and Ebola spread to understand how COVID-19 might behave.

SCREENING DRUGS –
Our supercomputers are allowing us to expedite testing, screen more than 8,000 drug compounds and found 77 have potential to fight against COVID-19. ... what took days on Summit would take months with a MacBook.

COORDINATING AND EXPANDING ACCESS FOR COVID-19 RESEARCH –
DOE made a nationwide call to the scientific community to utilize our state-of-the-art facilities and technologies to understand and combat COVID-19 together.

ENERGY.GOV

About

DOE User Facilities

NVBL Structure

NVBL Coordination Team

National Virtual Biotechnology Laboratory (NVBL)

<https://science.osti.gov/nvbl>

- Consortium of 17 DOE National Laboratories
- Takes advantage of DOE user facilities
- Initial activities include:
 - Epidemiological and logistical support
 - Addressing supply chain bottlenecks by harnessing advanced manufacturing
 - Medical therapeutics: computational drug discovery and structural biology
 - Innovations in testing capabilities
 - New project in understanding fate and transport of virus in the environment

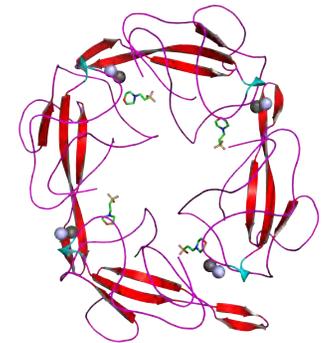
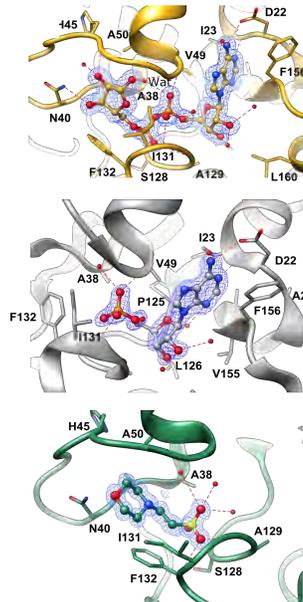
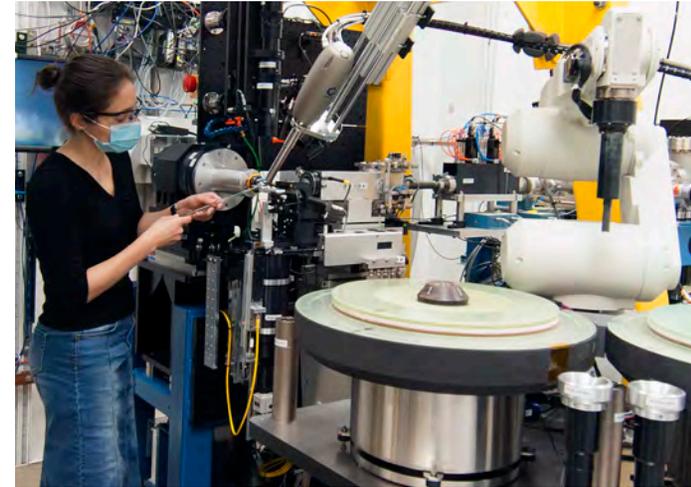
NVBL COVID-19 Task 5 Team

- **Structure-Based Protein Design for Diagnostics**

- Argonne National Laboratory
- Lawrence Berkeley National Laboratory
- Los Alamos National Laboratory
- Oak Ridge National Laboratory

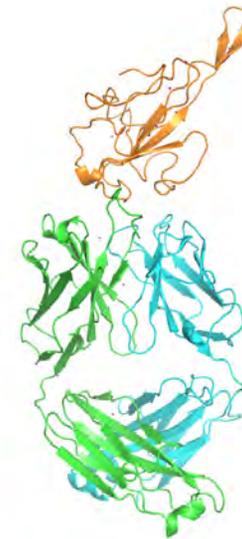
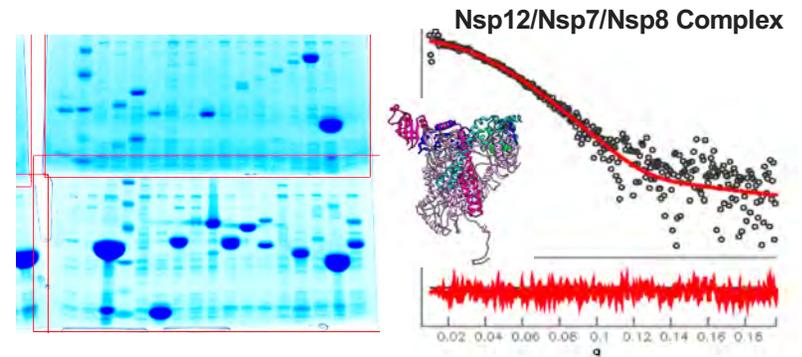
- **Structural Biology and Structure-based Drug Discovery**

- University of Chicago
- Northwestern University
- Argonne National Laboratory
- Auburn University
- University of Iowa
- Diamond Light Source
- University of Texas SWMC
- NIH/NIAMS



NVBL Structure-Based Protein Design for Diagnostics

- An integrated approach for utilizing emerging structural data has been applied to develop novel targets and demonstrate high-affinity reagents for non-nucleic acid-based detection systems.
 - The ANL team provided proteins for affinity reagent development for COVID-19 diagnostics.
 - The LBNL team performed HTP SAXS experiments at the ALS.
 - The ORNL team contributed to neutron protein crystallography studies and structure-based development of diagnostics.
 - LANL used protein design tools, available structures and HTP screening to develop affinity reagents for diagnostic tests.

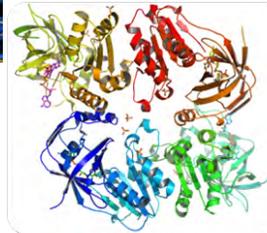
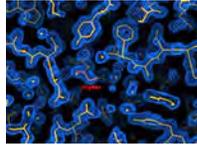
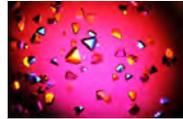


Advanced Photon Source Beamlines and APCF (APS Sector 84) Rapid COVID-19 Response



- The APS has stepped up as a world-leading source of information about SARS-CoV-2.
- More than 275 scientists from around the country have used 20 APS beamlines (remotely and mail-in) for nearly 7,700 hours, studying the virus with potential drugs, antibodies, and human proteins, and materials for better and less expensive N95 masks.
- 94 detailed structures for viral proteins, both alone and in complexes with potential therapeutic molecules, were determined using data collected at the APS.
- Argonne researchers have determined 40 of the structures identified using the APS.

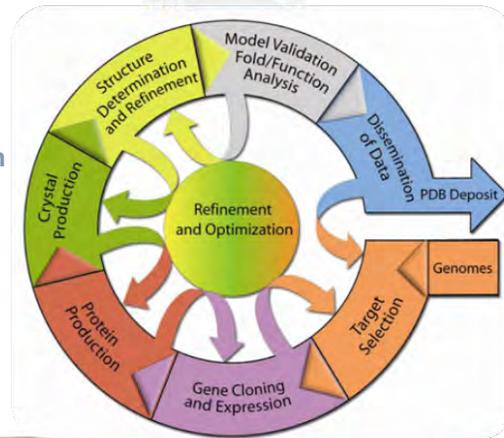
Argonne HTP Structural Biology Pipeline Applied to COVID-19



ATOM	1	N	MSE	A	1	0.189	14.507	14.365	1.00	48.72
ATOM	2	CA	MSE	A	1	0.485	13.096	13.983	1.00	49.75
ATOM	3	CB	MSE	A	1	1.995	12.409	13.909	1.00	51.19
ATOM	4	CG	MSE	A	1	2.316	11.290	13.878	1.00	57.21
ATOM	5	CE	MSE	A	1	4.222	10.700	13.827	0.90	72.87
ATOM	6	CE	MSE	A	1	5.166	12.464	13.847	1.00	69.67
ATOM	7	C	MSE	A	1	-0.228	12.091	14.925	1.00	47.56
ATOM	8	O	MSE	A	1	-0.026	12.116	16.153	1.00	47.90
ATOM	9	N	SER	A	2	-1.000	11.188	14.307	1.00	44.17
ATOM	10	CA	SER	A	2	-2.063	10.443	14.948	1.00	40.90
ATOM	11	CB	SER	A	2	-3.903	11.238	15.015	1.00	41.23
ATOM	12	CG	SER	A	2	-3.964	11.128	16.224	1.00	41.19
ATOM	13	C	SER	A	2	-2.394	9.275	14.045	1.00	38.41
ATOM	14	O	SER	A	2	-2.294	8.410	12.834	1.00	37.93
ATOM	15	N	PHE	A	3	-2.847	8.164	14.613	1.00	34.97
ATOM	16	CA	PHE	A	3	-3.080	6.993	13.831	1.00	32.04
ATOM	17	CB	PHE	A	3	-2.314	6.810	14.394	1.00	31.89
ATOM	18	CG	PHE	A	3	-2.481	4.538	13.604	1.00	30.34
ATOM	19	CD	PHE	A	3	-1.939	4.410	12.325	1.00	26.52
ATOM	20	CE1	PHE	A	3	-1.134	3.210	11.974	1.00	28.08



HTP crystallization
12 million drops to date, Mosquito, Robohotels

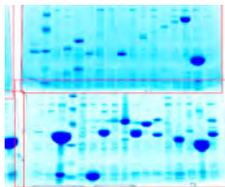


Robotic data collection and semi-automated structure determination

>3,300 PDB deposits to date (MCSG, SBC and CSGID) (4/week) SBC-Collect, HKL-3000, robotics

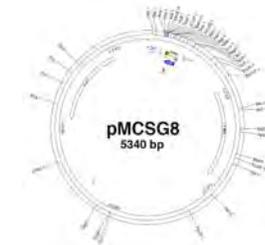


```
CGATGCGTGGTATTATTAGACA
ATACCGGAAAAGATATCTTTCTT
TTGAAAATGGCGAACTTTATTTG
TCAAAAGTCCATCTTTGGGCATCT
FATTGGATCAATGCGAGATATTTG
AAAATCCGAGATATGATCCCAAGC
CCAAAGTGGATATAAATGATAA
STAAGTTATAATCAACTCTTATG
ATTATGAAAAAGAACTAGAAAGC
AGACAATAATTCCTAGAGATGTA
FCTCTATTATGTGTTTCTCGGGTT
GGTCTTAATGGTAAAAATCTTTTG
FGGGTTTCATCAAAAGCTCTATTGC
FAAAACAAGAAGCTTTCCACAAA
ITTTTGGGAAAAAATGGTTAAA
```



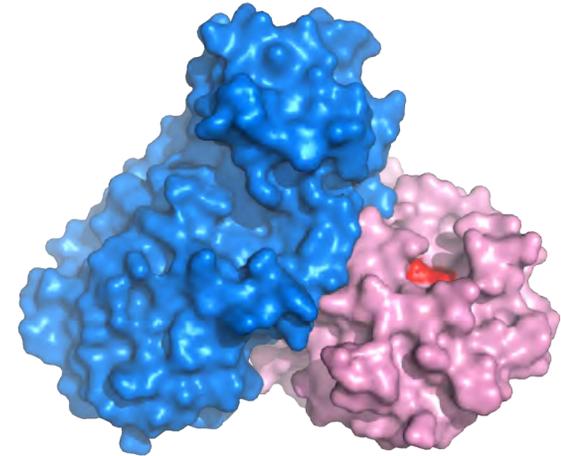
Mg-scale protein purification
>20,000 proteins to date
(max: 300/week)
AktaXpress systems

Cloning and small-scale protein expression
>65,000 clones to date (max: 4000/week)
>50 ug-scale protein production
(max: 4000/week) 250 genomes; >90 vectors; *E. coli*
and eukaryotic systems

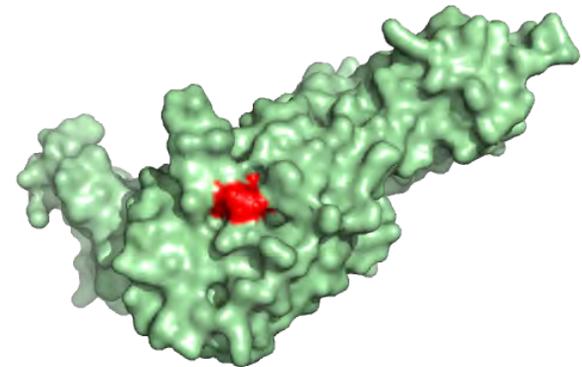


SARS CoV-2 Drug Targets

- Proteases are important SARS-CoV-2 enzymes potentially targetable with antivirals: papain-like protease (PLpro) and main protease (Mpro).
- Proteases are especially attractive targets because they play an essential role in several viral replication processes, including cleavage and maturation of viral polyproteins, assembly of the replicase-transcriptase complex, and disruption of host viral response machinery to facilitate viral proliferation and replication.
- PLpro and Mpro are conserved across different coronaviruses and promising inhibitors have already been discovered for their SARS-CoV-2 variant.]



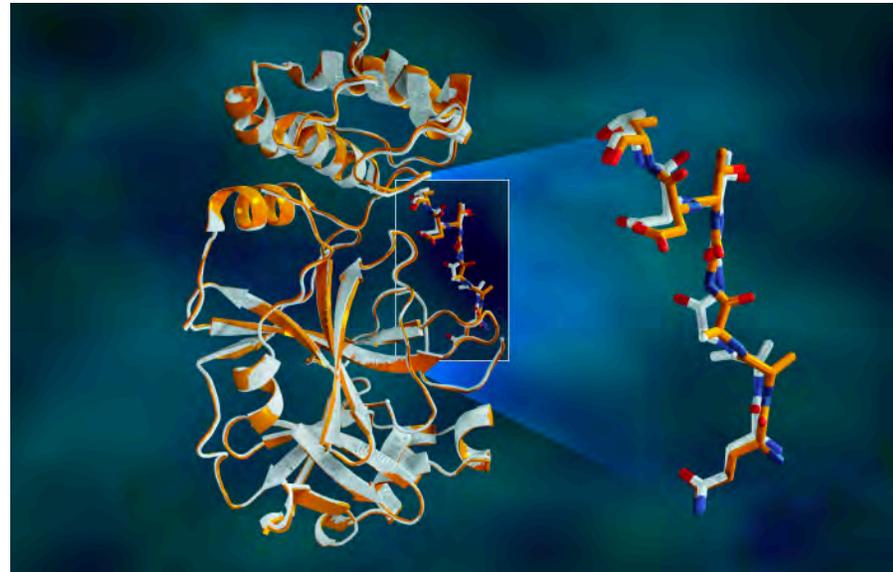
Structure of SAR-CoV-2 Mpro dimer



Structure of SAR-CoV-2 PLpro monomer

Structural Plasticity of SARS-CoV-2 3CL M^{pro} Active Site Cavity Revealed by Room Temperature X-ray Crystallography

- A team of researchers at the DOE's Oak Ridge and Argonne National Laboratories has performed the first room-temperature X-ray measurements on the SARS-CoV-2 main protease — the enzyme that enables the virus to reproduce.
- The model will be used to advance supercomputing simulations aimed at finding drug inhibitors to block the virus's replication mechanism. Research results are publicly available (PDB id: 6WQF) and have been published in the journal Nature Communications.



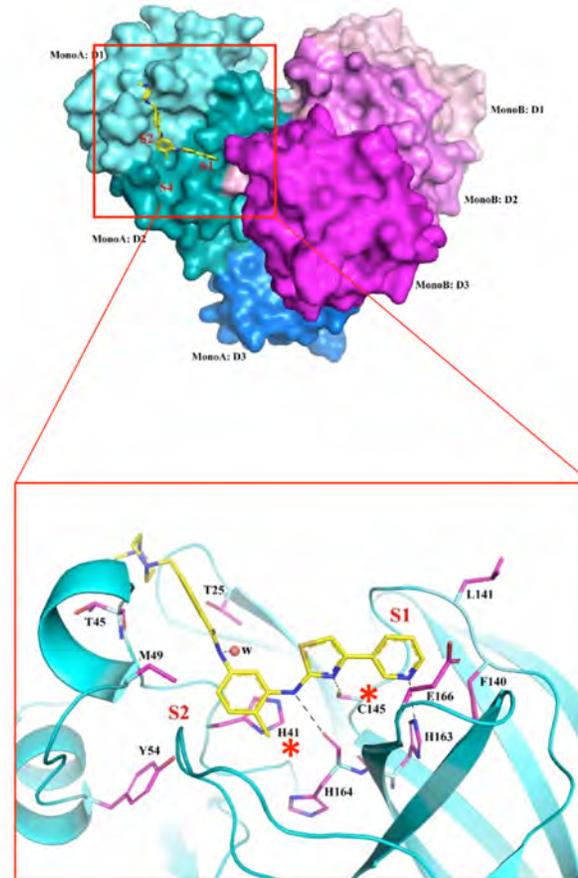
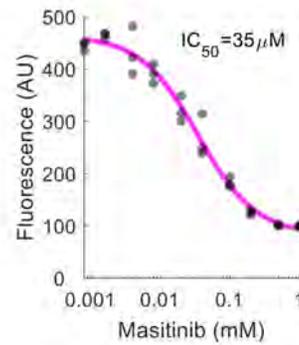
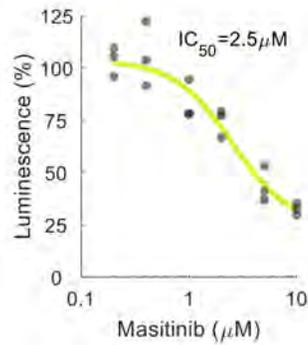
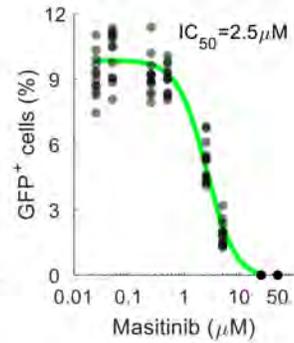
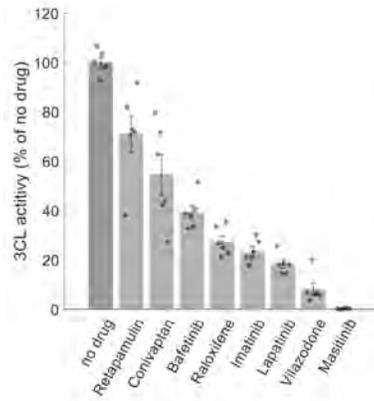
Overlapping X-ray data of the SARS-CoV-2 main protease shows structural differences between the protein at room temperature (orange) and the cryogenically frozen structure (white). Credit: Jill Hemman/ORNL, U.S. Dept. of Energy

17 structures determined by ANL and ORNL teams



Structural plasticity of SARS-CoV-2 3CL M^{pro} active site cavity revealed by room temperature X-ray crystallography. Kneller DW, Phillips G, O'Neill HM, Jedrzejczak R, Stols L, Langan P, Joachimiak A, Coates L, Kovalevsky A. Nature Communications volume 11, Article number: 3202 (2020)

Structure of SARS-CoV-2 Mpro with Masitinib

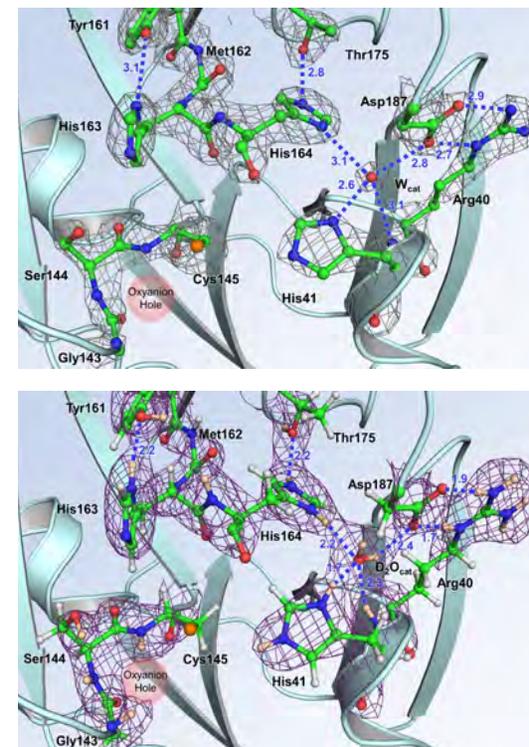


Room temperature X-ray and neutron crystallography of SARS-CoV-2 M^{pro} for the design of specific inhibitors

- Main Protease (3CL M^{pro}) is a major target for structure-guided drug design against SARS-CoV-2.
- The neutron structure shows that the catalytic site natively adopts a zwitterionic reactive state
 - His41 is doubly protonated and positively charged, whereas Cys145 is negatively charged in the thiolate state
- Combined X-ray and neutron studies play a critical role for *in silico* docking, clinical drug repurposing, and rational drug design efforts
- Scientific contributions
 - 5 papers published
 - 9 deposited X-ray structures reveal malleability of the active site, rare oxidation state, and binding mode clinical protease inhibitors



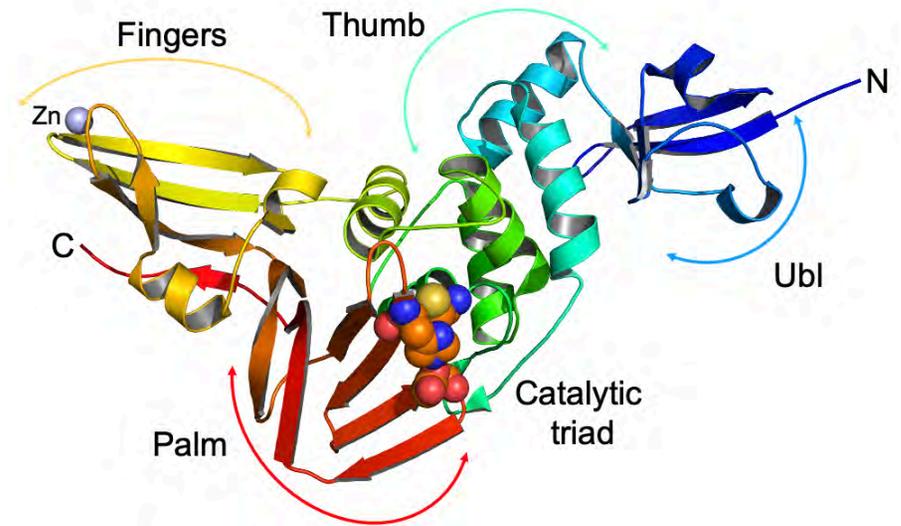
Catalytic site of SARS-CoV-2 3CL^{Mpro}



Top: Electron density map (no hydrogen atoms visible)
Bottom: Nuclear density map allowing visualization of protonation states and hydrogen bonding interactions

Structure of SARS-CoV-2 Nsp3 PLpro

- Cysteine PLpro protease is highly conserved and found in all coronaviruses, often in two copies
- PLpro exhibits multiple functions, in addition to processing polyproteins and recognizing LXGG sequence motif it has deubiquitinating activity and deISG15ylating (interferon-induced gene 15) activities
- Inhibitors block the peptidase activity of PLpro *in vitro* and some can block SARS-CoV-2 replication in cell culture assays.
- Our collection of structures provides fundamental molecular and mechanistic insight to PLpro and illustrates details for inhibitors recognition and interactions.

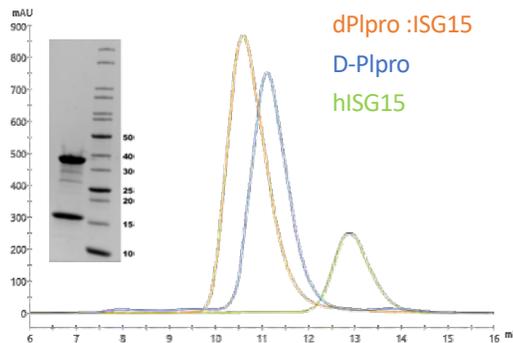


9 structures determined at ANL

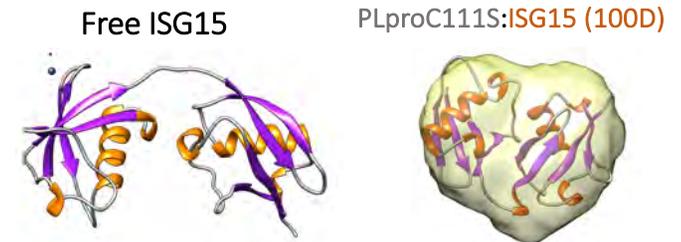
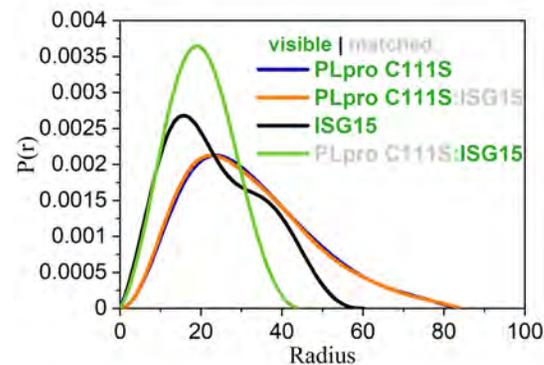
SANS provides insights into the deISGylating activity of PLpro and can be used to guide anti-viral drug design

- The PLpro plays important roles in viral polyprotein processing and deubiquitination and deISGylation of host proteins to counteract innate immunity
- The PLPro complex with human interferon-stimulated gene 15 (ISG15) is as an important structural target
- SANS with contrast matching showed an unexpected conformational rearrangement of ISG15 that was not observed in previously reported crystal structures of the complex

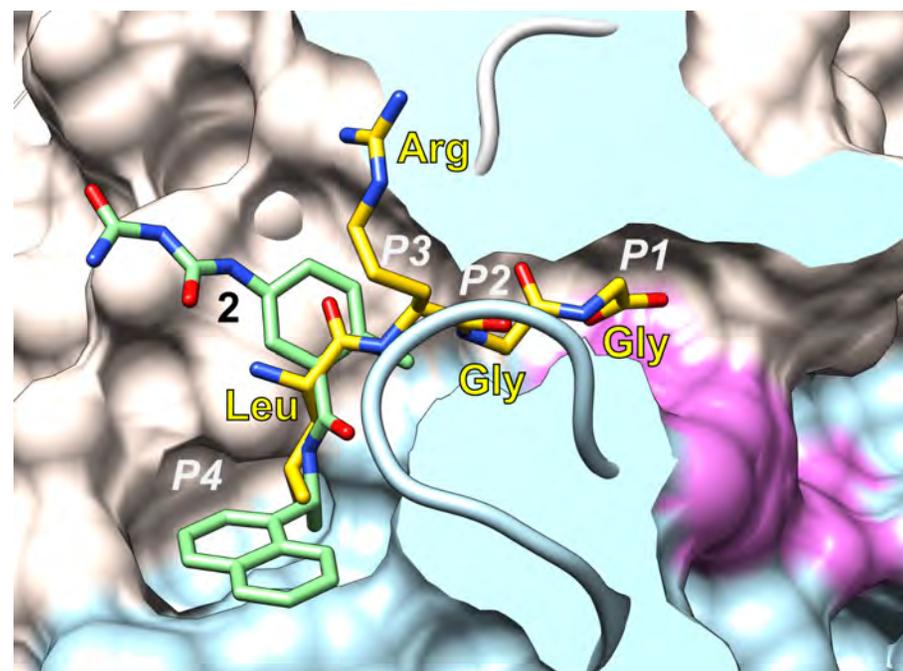
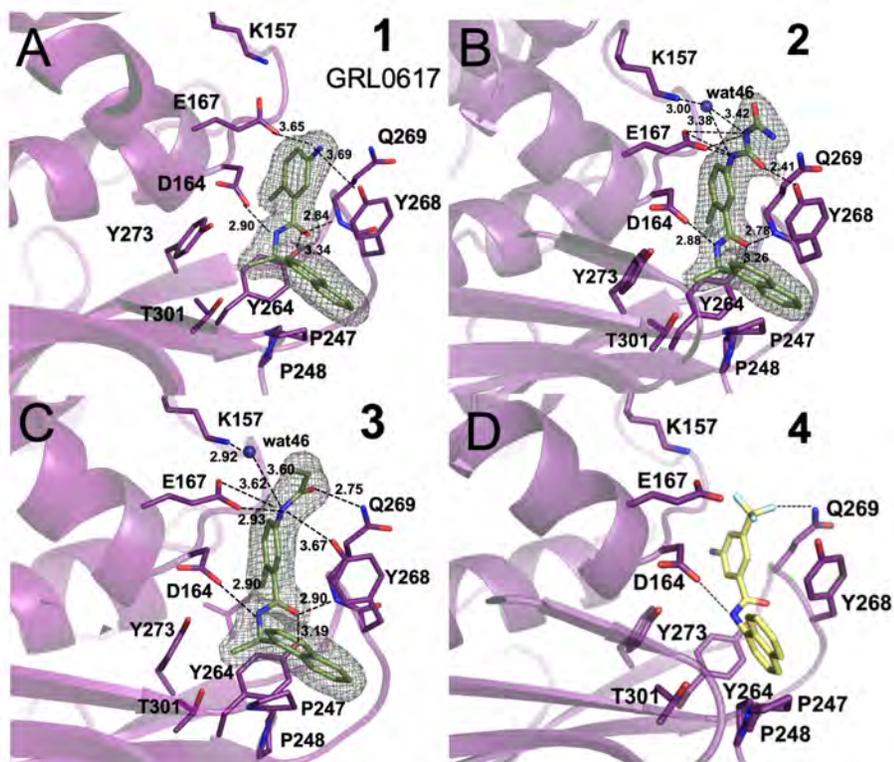
Size exclusion chromatography



Small-angle neutron scattering

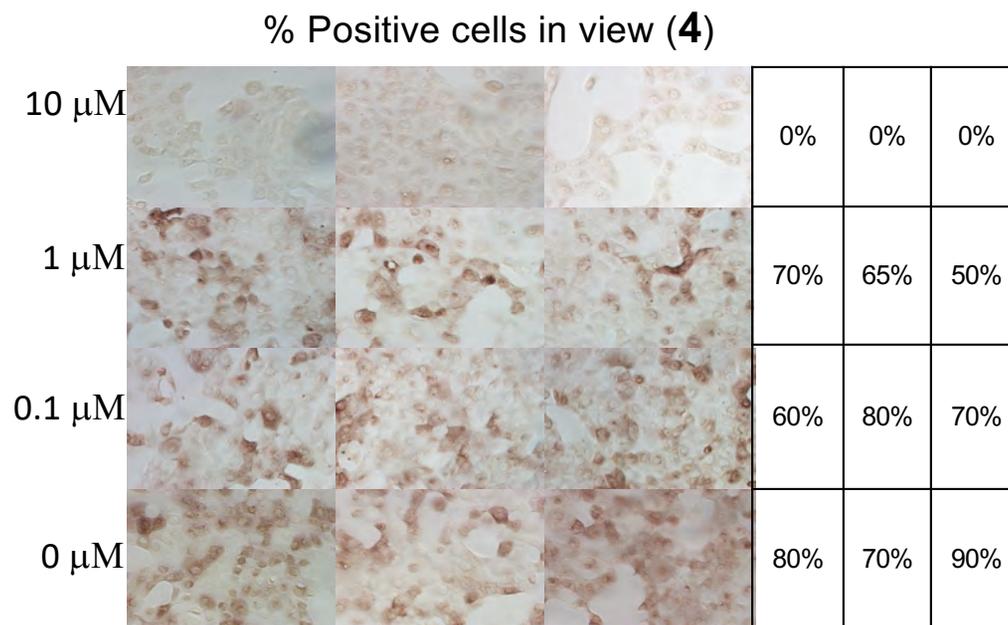
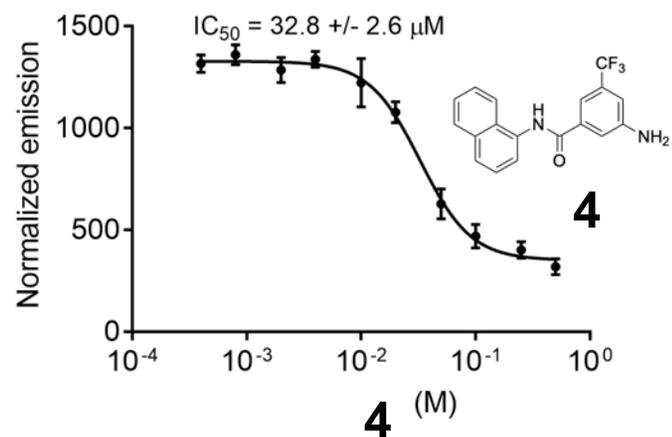
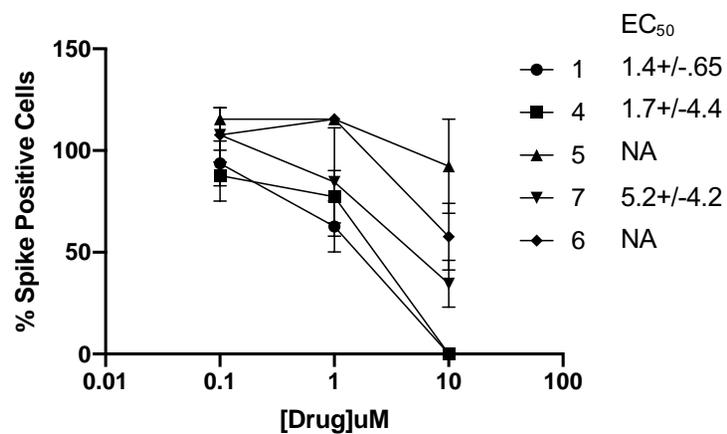


Structures of PLpro in Complex with Inhibitors Shows Details of Interactions



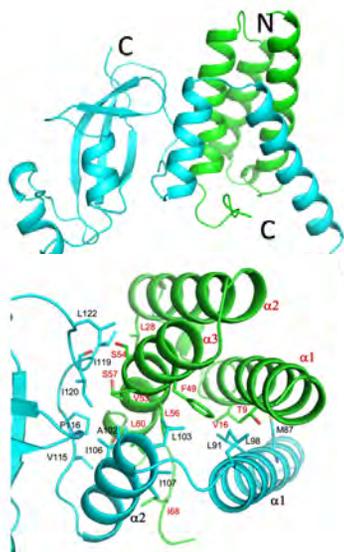
Inhibitor blocks access to PLpro active site

Compounds Targeting PLpro Inhibit SARS-CoV-2 Replication



Assembly, Mechanism and Inhibition of SARS-CoV-2 RNA Transcription Complex (Nsp7/8/12)

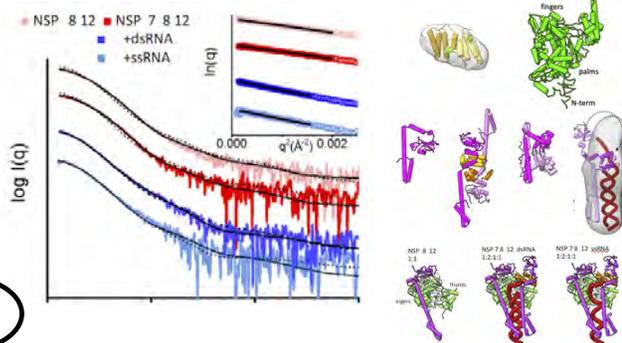
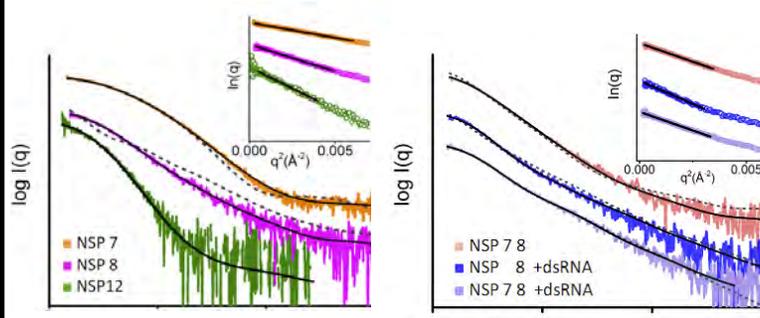
Argonne
NATIONAL LABORATORY
Crystallography & Protein
Expression for Atomic Detail



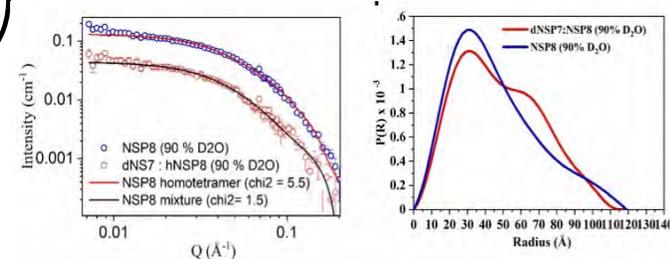
Nsp7/8 Crystal Structures



Solution Scattering SAXS for
High-Throughput Structures
Constrained by Crystallography

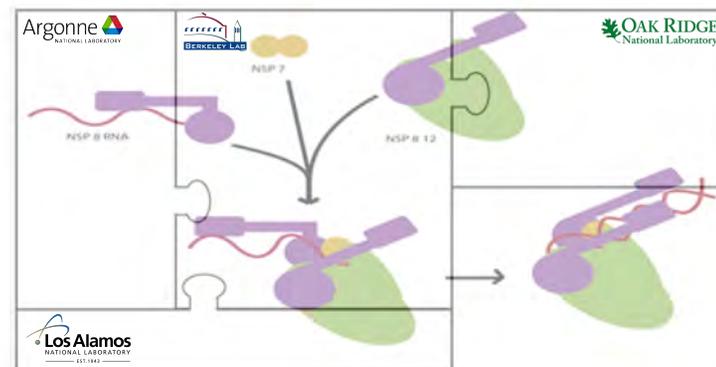


Contrast Matching SANS to
Visualize Conformational
Changes Induced During
Complex Formation



Los Alamos
NATIONAL LABORATORY
EST. 1943

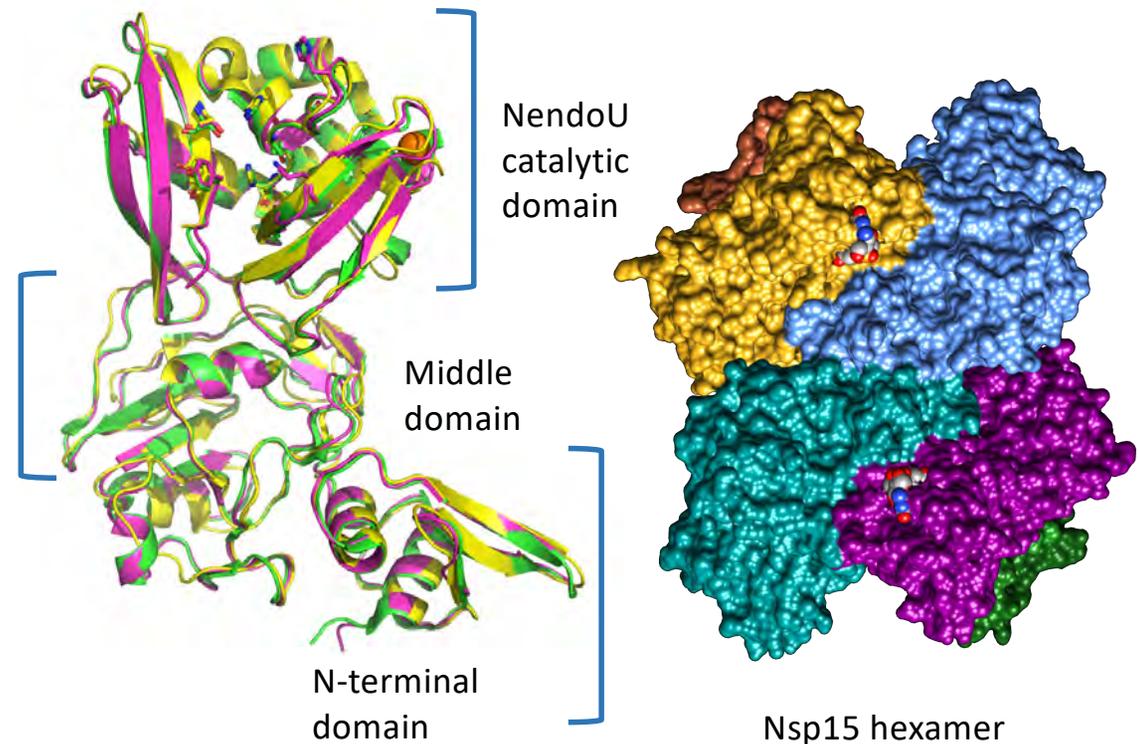
Computational Design of
Complex Disrupting Molecules



Integrated Data for Mechanism and Leads
on Inhibition

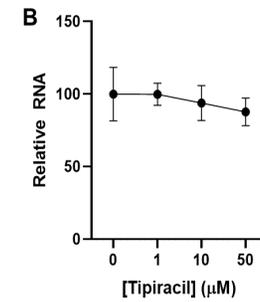
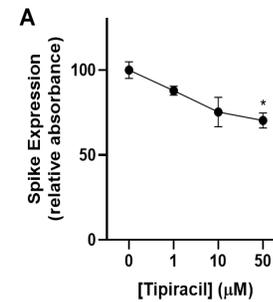
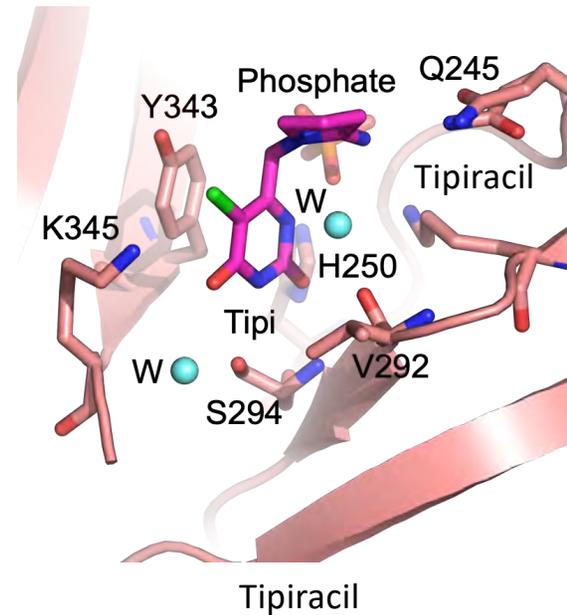
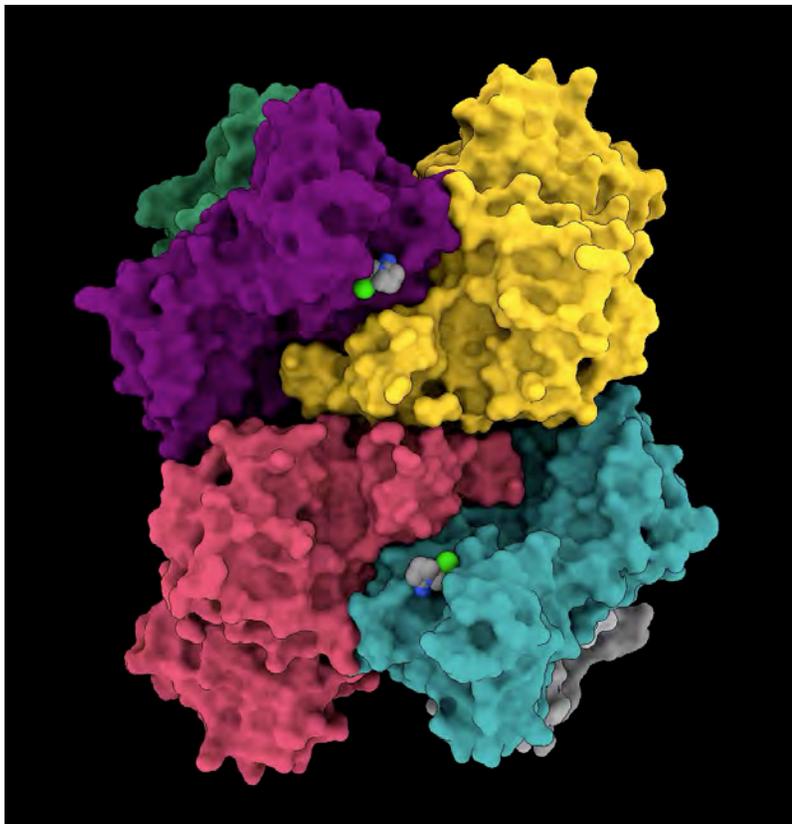
Nsp15 Endoribonuclease NendoU from SARS-CoV-2

- Nsp15 is a uridylate-specific endoribonuclease.
- NendoU activity interfere with the innate immune response and is essential in coronavirus biology.
- Nsp15 was shown to degrade viral RNA the polyuridine extensions on (-) sense strand of RNA.
- Nsp15 is highly conserved in coronaviruses.
- Several structures have been determined with ligands including complex with FDA approved drug Tipiracil, GpU product, 3'UMP Uridine Vanadate and 5'UMP.



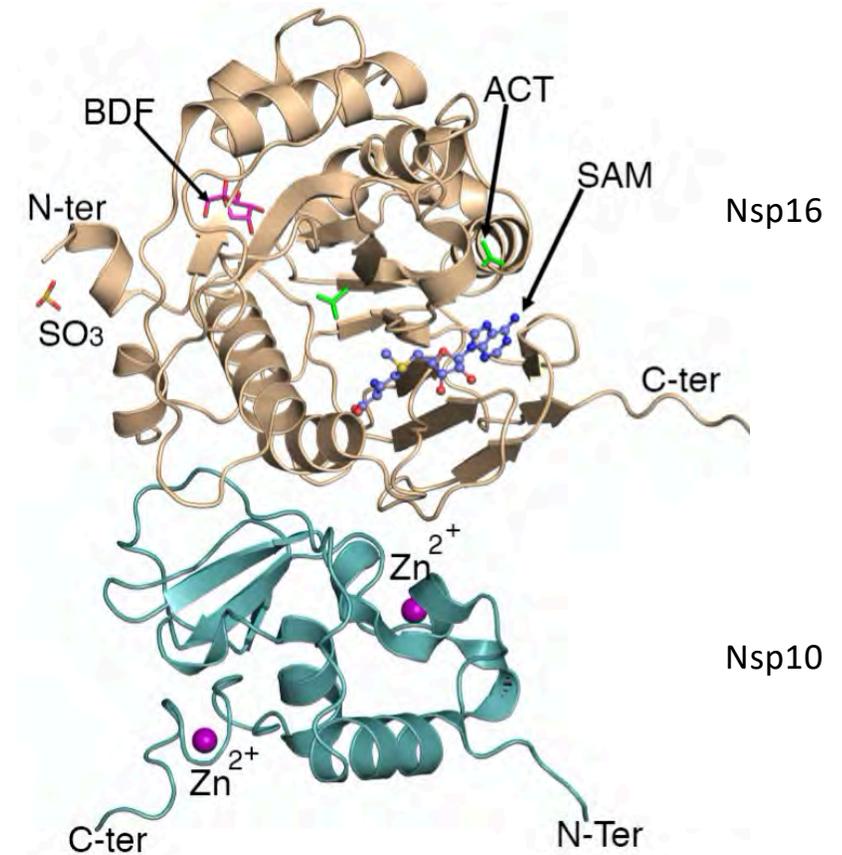
SARS-CoV-2 vs SARS-CoV-1 vs MERS-CoV

Nsp15 Endoribonuclease Complex with Tipiracil

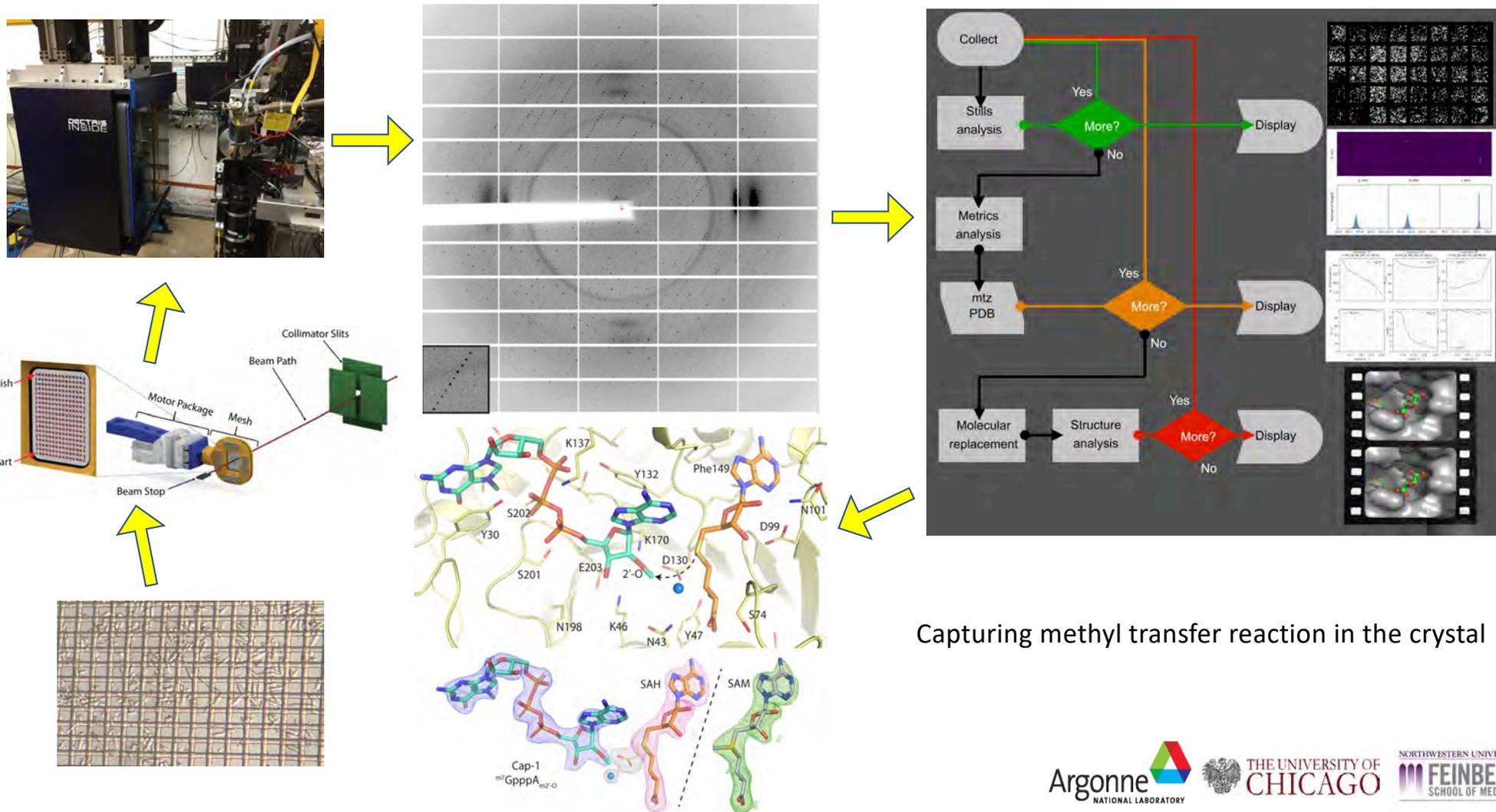


Nsp10/Nsp16 2'-O-methyltransferase from SARS-CoV-2

- Nsp10 and Nsp16 is responsible for the capping of mRNA at the 5' terminus which is critical for virus replication and fidelity.
- The methylation of the mRNA Cap is essential for efficient translation of viral transcript in a eukaryotic host.
- The presence of Cap-1 makes viral RNAs mimic the host transcripts and prevents their degradation.
- The active site of Nsp16 MTase is conserved in the coronavirus family; they utilize a K-D-K-E catalytic tetrad which is essential for enzymatic activity.



Serial Crystallography of SARS-CoV-2 Nsp10/Nsp16 Complex at 19ID

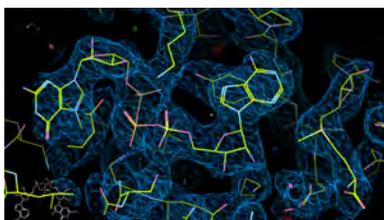


Capturing methyl transfer reaction in the crystal

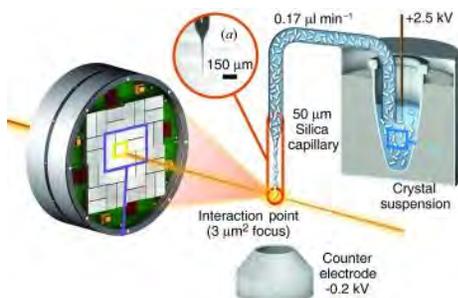
NVBL Partnerships in COVID-19 Research - Nsp10/Nsp16 and Mpro

Experiments at LCLS-MFX (SLAC) examine radiation sensitive Nsp10/16 structures and short-lived intermediates to provide new opportunities for the rational discovery of small molecule inhibitors.

- Damage free structures of Cap-1, the final stage in viral RNA maturation will verify serial synchrotron results.
- Visualization of acutely radiation sensitive catalytic metals (Mn or Mg) not observed in synchrotron data
- Capturing the rapid intermediate steps involved in docking of Cap-0 within Nsp10/16.



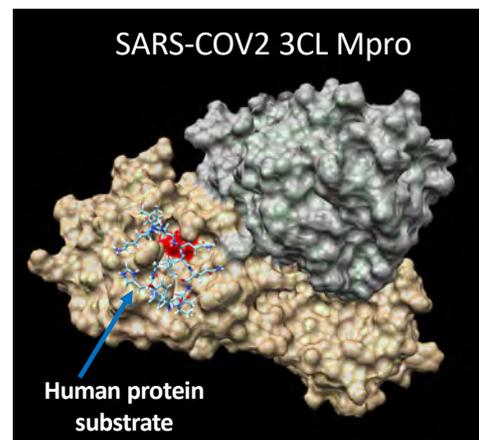
The SAM and m7GpppA ligands in the XFEL structure



The electro-spinning injector delivers tiny crystals into fs-scale X-ray pulses

A novel drug design concept against SARS-CoV-2 proteases

- 3CL Mpro interacts with human proteins affecting immune responses provoking side effects
- PLpro resembles human proteases (De-Ubiquitome)



Model of SARS-CoV-2 3CL Mpro with human protein substrates

- ❖ Extensive modeling and MD simulations
- ❖ Co-crystallization efforts ongoing
- ❖ SANS (ORNL), SAXS(SLAC-SSRL) planned

First experiments September/October, data analysis in progress. Fixed-target time-resolved measurements planned for 2021

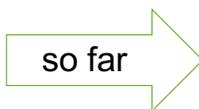
Novel inhibitor design concept for SARS-COV2 protease specificity, avoiding human-virus protein interactions

A Rapid Response at SLAC to Combat COVID-19

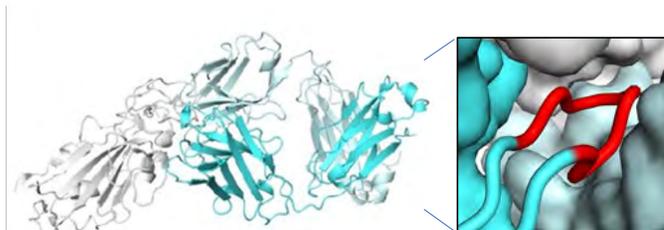


Fast Tracked Research Leads to Three Drugs in Clinical Trials + Two Entering Clinical Trials

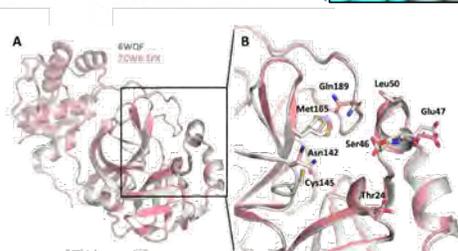
Synchrotron & CryoEM research started in March, LCLS in August



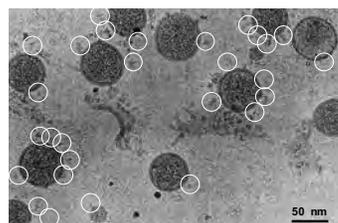
Proposals awarded time: **36** (5 international)
Fragments/inhibitors screened: **868**
PDB deposits: **18**



SSRL BL12-1: Human antibody (cyan) bound to the spike protein binding domain (grey). Close-up of the antibody and binding domain interface (Yuan, Science 2020)



LCLS-MFX: SARS-CoV-2 main protease structures at near-physiological temp to guide drug repurposing (Durdagi, bioRxiv/2020)



Cryo-EM: Structure of spike proteins and glycans of human coronavirus NL63 directly from virus particles (Zhang, bioRxiv/2020/245696)



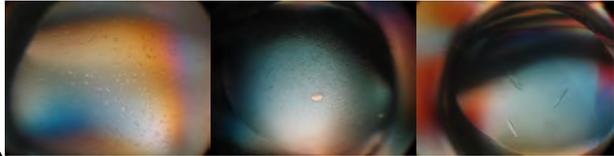
- Hamburg, Germany
- Oxfordshire, UK
- Istanbul, Turkey

Groups from across the US and abroad used SSRL, LCLS and CryoEM facilities at SLAC for COVID-19 related research

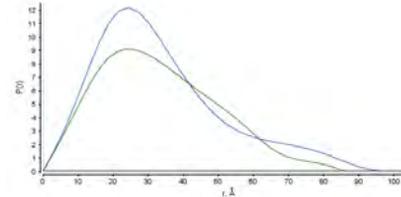
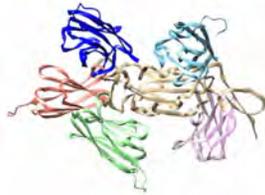
Diagnostic and Blocking Antibodies and Nanobodies Against Spike and Nucleocapsid

Nanobodies Against Spike (RBD)

Nanobody Production:
8/week – DNA to Crystal Trials



Using SAXS to test simultaneous binding by multiple nanobodies and conformational change to RBD



Goals & Applications

Enhanced Blocking



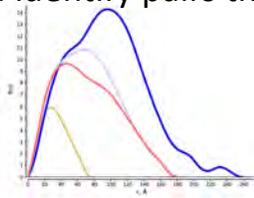
Antigen Detection



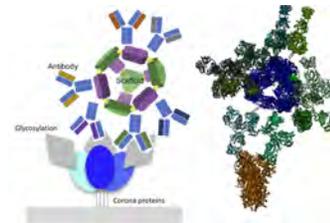
Antibodies Against Spike (RBD)

Using SAXS to identify variation in modes of binding and identify pairs that can bind simultaneously

Using designed proteins to display multiple antibodies for increased avidity



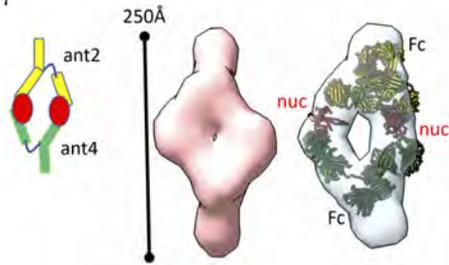
RBD
7,8,9_pool_RBD two antibody
7,8,9_pool_RBD one antibody
10,11_pool_RBD two antibody different



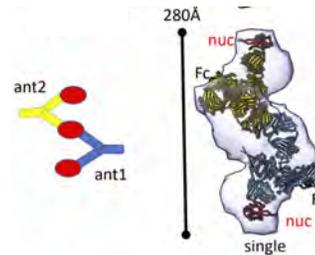
Hybrid Antibody Nanobody Systems



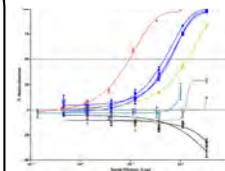
Antibodies Against Nucleocapsid



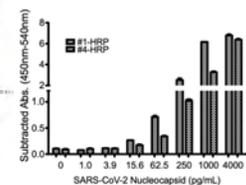
Using SAXS to distinguish modes of binding by pairs with and without detergents



Neutralization Assays



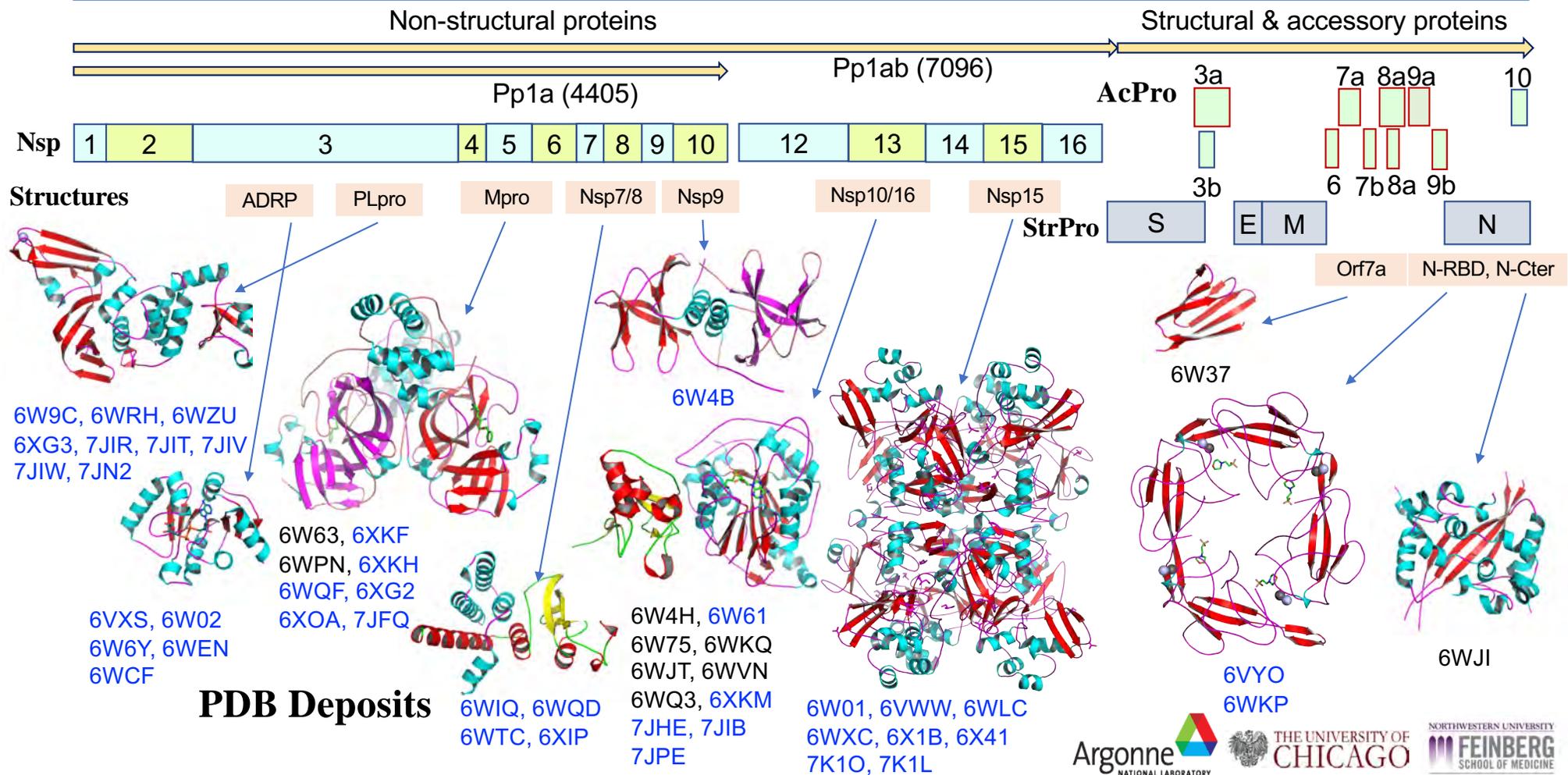
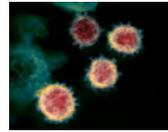
ELISA



Have systems with 100% blocking and pg/mL detection

SARS-CoV-2 Genome and CSGID (57) /UoC/ANL (40) Structures

(+) RNA (29903)



Acknowledgements

ANL/UoC/CSGID

C. Chang
G. Babnigg
M. Endres
R. Jedrzejczak
Yc. Kim
N. Maltseva
K. Michalska
J. Osipiuk
L. Stols
K. Tan
C. Tesar
M. Wilamowski



Univ. of Chicago

A. Baranczak
B. Dickinson
G. Dong
N. Drayman
T. Kossiakoff
G. Randall
S. Snyder
J. Solway
S. Tay
P. Wilson

BioCARS/UoC

V. Sraayer

Auburn Univ.

J. Wower

CSGID

D. Borek, UT, SWMC
D. Fremont, WashU
A. Godzik, UCR
R. Kuhn, PU
Z. Otwinowski, UT, SWMC
W. Minor, UVA
A. Mesecar, PU
K. Satcher, NU
A. Savchenko, UC

SLAC/LCLS/SSRL

S. Boutet
A. Cohen
M. Hunter
A. Lyubimov
R. Sierra
S. Wakatsuki
J. Wierman
The construction of the MFX instrument at LCLS was BER funded and operates as a partnership between LCLS and SSRL-SMB

ORNL

H. O'Neil
L. Coates
A. Kovalevsky
J. Parks

LANL

R. Jha

ALS/LBNL/SIBYLS

P. Adams
G. Hura
M. Hammel
C. Hodge
D. Rosenberg
S. Tsutakawa

Oxford/Diamond

F. von Delft
M. Walsh



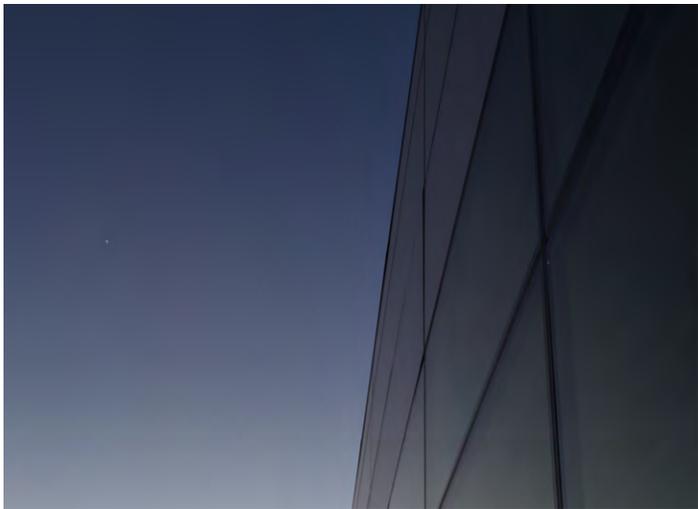
ANL/CELS

R. Stevens
A. Ramanathan

SBC, ANL/APS/XSD

D. Sherrell
A. Lavens
K. Lazarski

Funding is provided by: the National Institute of Allergy and Infectious Diseases of NIH under Contract No. HHSN272201700060C, the DOE Office of Science through the National Virtual Biotechnology Laboratory, a consortium of DOE national laboratories focused on response to COVID-19 (the Coronavirus CARES Act) and the use of Structural Biology Center beamlines at the Advanced Photon Source is supported by the U.S. DOE Office of Science and operated for the DOE Office of Science by Argonne National Laboratory under Contract No. DE-AC02-06CH11357.



Thank you
